## Review paper

# Mitomycin C: mechanism of action, usefulness and limitations

## Jaap Verweij<sup>CA</sup> and Herbert M. Pinedo

J Verweij is at the Department of Medical Oncology, Rotterdam Cancer Institute, Daniel den Hoed Kliniek, Groene Hilledijk 301, 3075 EA Rotterdam, The Netherlands. HM Pinedo is at the Department of Medical Oncology, The Netherlands Cancer Institute and Free University Hospital, Plesmanlaan 121, 1066 CX Amsterdam, The Netherlands.

The mitomycins are antitumor antibiotics that are under investigation now for more than 30 years. Mitomycin C (MMC) is the best investigated subtype. It serves as a prototype for drugs with bioreductive alkylation, which is a unique feature of this class. MMC is mainly active under anaerobic circumstances. The pharmacokinetics are linear in a two-compartment model. The main toxicities of MMC are thrombocytopenia and leucocytopenia. Rare but severe side effects are a hemolytic uremic syndrome, pneumonitis and cardiac failure. MMC has a wide clinical antitumor spectrum with efficacy in various tumor types such as gastric cancer, pancreatic cancer, breast cancer, non-small cell lung cancer, cervical cancer, prostate cancer and bladder cancer. Still, the above mentioned side effects prevent a more widespread use. The most important features of the drug will be reviewed.

Key words: Mitomycin C, bioreductive alkylation, clinical pharmacology, hemolytic uremic syndrome.

### Introduction

The mitomycins constitute a class of antitumor antibiotics isolated from *Streptomyces caespitosus*. Mitomycin C (MMC), isolated in 1958,<sup>1</sup> has received the greatest attention of all mitomycins, both in preclinical and clinical investigations. The drug has a wide spectrum of antitumor activity, but clinically also exerts infrequent but sometimes quite severe side effects that prevent a more widespread use.

#### Chemistry and mechanism of action

MMC was isolated from fermentation filtrates of Streptomyces caespitosus as blue violet crystals. The

crystal structure and the absolute stereochemical configuration of MMC have been determined.<sup>2</sup> MMC has a molecular weight of 334 daltons and is soluble in water and organic solvents. Its structure is shown in Figure 1.

The structure, in which quinone, aziridine and carbamate functions are arranged around a pyrrolo[1,2-a] indole nucleus, is quite unique. Under acidic conditions protonation of the  $C_{9a}$ -methoxy group triggers cleavage<sup>3</sup> resulting in a  $C_9$ - $C_{9a}$  double bond and opening of the aziridine ring<sup>4</sup> (Figure 2), while additional replacement of the  $C_7$ -amino function by a hydroxyl group and cleavage of the  $C_{10}$ -carbamate function occur at more pronounced acid conditions.<sup>5</sup> Under alkaline conditions the latter two changes are seen<sup>6,7</sup> without concomitant effects on the  $C_{9a}$ -methoxy group or the aziridine ring.

In principle, MMC inhibits DNA synthesis. Oxidized MMC is inactive: the drug requires chemical or enzymatic reductive activation to either the corresponding semiquinone or the hydroquinone form, to bind DNA by mono- or bifunctional

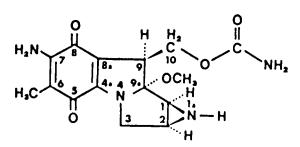


Figure 1. Structure of mitomycin C.

CA Corresponding Author

Figure 2. Mechanism of acid-catalyzed hydrolysis of mitomycin C (R = H).

alkylation.<sup>8</sup> Bifunctional alkylation is thought to lead to crosslinking of strands of double helical DNA.<sup>9</sup> A second DNA alkylation promoting mechanism, which appears to be less important, is acidic activation.<sup>10,11</sup>

The reduction of MMC which is necessary for alkylating reactions has led to the introduction of the term 'bioreductive alkylation' to describe the mechanism of action.<sup>12</sup> MMC is considered to be the prototype of the bioreductive alkylating agents.

Under anaerobic conditions a one- or two-electron reduction with subsequent spontaneous loss of methanol leads to the formation of an unstable reactive intermediate. The formation of a quinonemethide<sup>13</sup> then results from rearrangement of the hydroquinone followed by a nucleophilic addition of DNA leading to a mono-alkylated product.14 Intramolecular displacement of the carbamate group would then result in the cross-linked adduct (Figure 3). Gradual addition of a reducing agent<sup>15</sup> or excess addition16 results in a higher binding frequency. These conditions are favorable for the maintenance of the semiquinone radical, the intermediate which is formed by the first electron uptake of MMC. The semiquinone is therefore believed to initially bind to DNA. The existence of this radical during the process of MMC reduction has been proven and evidence is increasing that one-electron

reduction is sufficient to activate both the  $C_1$  and  $C_{10}$  electrophilic centers.

The fate of MMC after reductive metabolization under aerobic conditions is different. Either the

Figure 3. Reductive activation and DNA alkylation of mitomycin C.

semiquinone radical<sup>17-21</sup> or the hydroquinone reacts with molecular oxygen to generate superoxide radical anions, 17-21 hydroxyl radicals 22,23 or hydrogen peroxide.24 Cytotoxicity of these highly reactive forms may be exerted through lipid peroxidation or nucleic acid damage, which can in turn be prevented by free radical scavengers such as mannitol as well as by protective enzymes such as superoxide dismutase or catalase. The type of reactive MMC intermediate is a result of the half-life of the semiquinone radical: in an aprotic environment this is the semiquinone itself due to a long half-life; in protic media the semiquinone exists only a few milliseconds before rapid uptake of a second electron results in the formation of the hydroquinone. Besides, O<sub>2</sub> will play a role because it inactivates the semiquinone and this inhibits a further reduction.

The reductive activation of MMC can be initiated or modulated by enzyme systems such as DT-diaphorase, <sup>25,26</sup> NADPH-cytochrome P-450 reductase, <sup>27</sup> NADPH-cytochrome C reductase, <sup>28</sup> xanthine-oxidase <sup>28</sup> and some flavoprotein transhydrogenases, <sup>29</sup> and also by chemical reducing agents such as sodium thiosulfate <sup>15,30</sup> and sodium borohydride. <sup>30</sup>

Our knowledge on the process of alkylation of DNA nucleotides by MMC has been extended by mimicking the *in vivo* process by *in vitro* work with model nucleophiles in aqueous solution under reducing condition.<sup>31,32</sup> Potassium ethyl xanthate<sup>30</sup> and potassium ethyl monothiocarbonate<sup>32</sup> appeared to be ideal nucleophiles. Both mono- and disubstituted mitosenes with nucleophiles at C<sub>1</sub> and/or C<sub>10</sub> of MMC were identified.

By decreasing the temperature during the alkylation reaction it is possible to discriminate between these two reaction sites. <sup>32</sup> Such mild conditions lead to preferential substitution at  $C_1$ , while under more reductive conditions displacement of the carbamate group at  $C_{10}$  in the monofunctionally alkylated molecule is readily observed, <sup>32</sup> which implies that after monofunctional alkylation of MMC at  $C_1$ , secondary activation of monofunctionally bound MMC at  $C_{10}$  is probably the main route for the bifunctional binding of MMC to nucleophiles, and for cross-linking of DNA. <sup>20,26</sup>

Studies on covalent interactions between MMC and DNA or DNA fragments characterized adducts as mono- or bifunctional mitosene derivatives, similar to the observations with model nucleophiles. The binding sites of MMC in DNA are the N<sub>6</sub> position of adenine residues or either the N<sub>2</sub> or N<sub>7</sub> position of guanine residues. 9,11,33,34 The formation of O<sup>6</sup>-guanine-MMC adducts may also be impor-

tant.<sup>34,35</sup> Acid-activated MMC was found to alkylate preferentially the guanine N<sub>7</sub> position, in contrast to reductively activated MMC, which preferentially alkylates the guanine N<sub>2</sub> position,<sup>11</sup> possibly by the different electronic structures of acid- and reduction-activated MMC. Evaluation of the activation mechanism of MMC can presumably now be deduced from analysis of the DNA adducts formed in vivo.

Activated MMC is most effective in the late  $G_1$ and S phases of the cell cycle.<sup>36</sup> MMC does not reduce the size of nucleotide pools<sup>37</sup> and has no effect on DNA polymerase from HeLa cells.36 DNase activity has not been observed.<sup>38</sup> Consequently, besides the specific inhibition of DNA synthesis secondary to alkylation, inhibition of both RNA synthesis inhibition and protein synthesis inhibition seems to be non-specific manifestations of cell toxicity. 39,40 There are several indications that DNA repair is not inhibited by MMC. 41-43 MMC has been shown to produce chromosomal abnormalities, such as chromatid exchanges in leukocytes, but whether these are related to mitotic inhibition, or inhibition of DNA synthesis, is unclear.<sup>36</sup> MMC is teratogenic<sup>36</sup> as it increases the incidence of tail and paw abnormalities, as well as the incidence of spontaneous abortions, in rats and mice. MMC is also carcinogenic. 44,45

Although a definitive proof is still lacking, there is strong evidence that MMC is preferentially activated to a cytotoxic intermediate in hypoxic environments. 46 50 In vitro a preferential activation of MMC to cytotoxic metabolites in hypoxic tumor cells has been demonstrated in the EMT-6 and S-180 murine cell lines.<sup>23</sup> On the other hand, such selective toxicity could not be observed in vivo. 46,47 However, recently it was shown that dicoumarol, an inhibitor of DT-diaphorase, increases the cytotoxicity of MMC to hypoxic EMT-6 cells in vitro and in vivo, 51 possibly by inhibition of enzymes involved in the activation and inactivation of MMC. The increase in cytotoxicity to hypoxic tumor cells did not coincide with an increase of toxicity to well-oxygenated tissues such as the bone marrow.<sup>51</sup> In view of this preferential toxicity for hypoxic cells, a combined regimen of MMC and radiation is considered interesting for the treatment of solid tumors, because radiation will be more effective to aerobic tumor cells.

The mechanism(s) of resistance to MMC is not completely understood but probably involves changes in drug accumulation, bioactivation of the alkylating species, and DNA excision repair.<sup>52</sup> In Chinese hamster ovary cell mutants increasing

drug-resistance was related to a progressive loss of MMC activation capacity and increasing capacity for excision repair of DNA. The specific bioactivation enzyme system deficient in these resistant cells has not yet been identified. Also MMC appears to share in the multidrug resistance phenotype that encompasses doxorubicin, vincristine and other natural products, and appears to be mediated by amplification of the P-170 transmembrane drug efflux glycoprotein.

## Clinical pharmacology

Extensive pharmacokinetic data on MMC have become available since the introduction of different modifications of a high performance liquid chromatography (HPLC) assay. 53-58 All these assays have a detection limit of approximately 1 ng/ml sample. There is a biexponential decline of the plasma concentration time curves, indicating a two-compartment model with linear pharmacokinetics up to doses as high as 60 mg/m<sup>2</sup>. After a rapid half-life of distribution (2-10 min), the elimination half-life is 25-90 min (mean 54 min). No correlations have been found between pharmacokinetic data of MMC and a wide variety of clinical parameters. 53-55 Most importantly, impaired liver and renal function do not appear to change the pharmacokinetic behavior of MMC and therefore do not require dose reductions. Two studies reported an unexplained increased total body clearance and decreased area under the plasma concentration time curve of MMC following combination chemotherapy also including 5-fluorouracil and doxorubicin.<sup>54,59</sup>

Because urinary recovery after i.v. administration ranged from 1-20%, which cannot explain the rapid plasma clearance, it has been suggested that MMC is rapidly cleared from plasma by biodegradation. The liver is thought to be the major organ of biotransformation but the spleen, kidney, brain and heart may also be involved in the process. 60,61 The presence of oxygen markedly reduced the rate of metabolism of MMC in liver homogenates, as compared to the metabolism in a similar anaerobic system. 60 As biotransformation is required for activity, this supports the theory of a more pronounced activity under anaerobic conditions. After intraarterial hepatic infusions, only a three-fold greater regional exposure was found,56 which in view of the hepatic extraction of MMC, which is only about 20%, suggests a very limited benefit of this technique with respect to reduced systemic toxicity. A variety of microspheres have been used in an attempt to increase local exposure to MMC and although they resulted in a reduced systemic exposure with a decrease in systemic peak levels and AUC<sup>62-65</sup> the increase in tumor exposure was insufficient to enhance tumor regression.<sup>62</sup>

Both the intraperitoneal<sup>66-68</sup> and intravesical<sup>69</sup> route of administration result in a significant local exposure advantage and very low plasma levels. While intravesical administration has been successful in treating bladder cancer, the precise role of intraperitoneal administration has yet to be delineated.

Although MMC is absorbed after oral administration, absorption has been shown to be rather erratic by this route. The MMC is usually administered as a 10–15 mg/m² bolus i.v. injection once every 6 weeks, because more frequent administration will result in severe bone marrow toxicity.

## **Toxicity**

The most frequent side effect of MMC is a delayed myelosuppression, which appears to be directly related to schedule and total dose. Thrombocytopenia is more frequent than leucocytopenia and anemia. Other toxicities include usually mild and infrequent anorexia, nausea, vomiting and diarrhea. Alopecia, stomatitis and rashes also occur infrequently. Extravasation results in tissue necrosis with very disabling ulcers that may require plastic surgery. Extremely high doses of MMC (60 mg per dose) may result in lethal veno-occlusive liver disease. Other infrequent, but potentially lethal, side effects include hemolytic uremic syndrome, interstitial pneumonitis and cardiac failure.

The pathogenesis of the MMC-induced hemolytic uremic syndrome<sup>75</sup> still remains unclear, although prostacycline deficiency may play a role.<sup>76</sup> The incidence appears to be less than 10%<sup>75,77</sup> and was suggested to be dose-dependent.<sup>77</sup> There is no consistently effective treatment for this syndrome.

Pulmonary toxicity of MMC consists of an interstitial pneumonitis. <sup>75</sup> Discontinuation of MMC administration may occasionally lead to recovery from this side effect, but usually there will be a progressive respiratory failure. Corticosteroid treatment may be helpful in preventing progression of pulmonary dysfunction. The incidence of pulmonary toxicity is approximately 7% of the treated population; cardiac failure secondary to MMC occurs in a similar percentage of treated patients, and rises with cumulative doses above 30 mg/m<sup>2</sup>. <sup>78,79</sup>

## Clinical antitumor activity

MMC has been studied extensively in advanced neoplasms in humans. The initial studies reported on the use of daily low dose schedules, which resulted in unacceptably severe and cumulative myelosuppression. For this reason, an intermittent dosing schedule was introduced, using bolus injections every 4–8 weeks, resulting in manageable hematologic toxicity. The following review will only refer to studies using the intermittent schedule (Table 1).

Single agent activity in gastric cancer was reported to be 29%. 80-82 Combination chemotherapy incorporating MMC achieves slightly higher response rates and also improves survival in the responding patients. 83-85

Single agent treatment with MMC in pancreatic cancer achieves a response rate of 27%. 86 This indicates that MMC is one of the most active drugs against this neoplasm presently available. Unfortunately, combination chemotherapy does not appear to improve these results. 84,86–92

Single agent activity in 293 breast cancer patients treated with MMC was 20%, 93-96 with a wide variation in response rates in different studies (0-31%), as a result of the fact that MMC was used either as first-, second- or third-line treatment. Recent studies indicate that second- and third-line treatment with MMC are far from rewarding. 96 In this tumor type, combination chemotherapy including MMC was found to be superior to single agent treatment with MMC, but response rates in larger series remain lower compared to more active combinations such as cyclophosphamide/methotrexate/fluorouracil (CMF) or fluorouracil/doxorubicin/ cyclophosphamide (FAC).

Treatment with MMC as a single agent in 148 patients has resulted in an overall response rate of 28% in non-small-cell lung cancer (NSCLC). 103-106 Data on survival are frequently not indicated. Data

Table 1. Single agent activity of mitomycin C

Tumor type	No. of evaluable patients	Response rate (%)
Gastric cancer	343	29
Pancreatic cancer	44	27
Breast cancer	293	20
Non-small-cell lung		
cancer	148	28
Cervical cancer	173	36
Colorectal cancer	272	16
Prostatic cancer	30	36

on combination chemotherapy with MMC included are quite inconsistent, with response rates varying from 20-59%. <sup>107–112</sup> However, as for other drug combinations, responses are usually short and survival benefit is not obtained.

In squamous cell cancer of the uterine cervix, single agent MMC treatment achieved an overall response rate of 36% in 173 patients. 113,114 Combination chemotherapy including MMC yields even higher responses. However the most important drug in this disease appears to be cisplatin and the decision as to whether to add other drugs such as MMC to cisplatin is presently under investigation in an ongoing European Organization for Research and Treatment of Cancer (EORTC) study. Survival for complete responders to combination chemotherapy is 12–30 months and for partial responders 6–15 months, both compared to a median survival of 3–6 months for progressive disease, which at least suggests a benefit.

In superficial bladder cancer intravesical instillations of 30–40 mg MMC in 20–40 ml sterile water and retained in the bladder over 2–3 hours have achieved an overall response of 67% in 276 patients 115–118, approximately two-thirds of them achieving a complete remission of long duration up to 25 months or more. Epodyl, thiotepa, and doxorubicin appear to be equally active in destroying superficial bladder tumors, but MMC probably is the least toxic. Data on i.v. treatment in more advanced disease are anecdotal.

MMC is one of the few agents with very modest activity against colorectal cancer. In a compilation of 272 patients, 44 responders were found (16%) with single agent treatment. In Consistent data on combination chemotherapy with MMC show only minor improvement of results compared to single agent treatment. All treatments including MMC do not create any survival benefit for responding patients. For this reason, it is not recommended to apply MMC in colorectal cancer.

In prostatic cancer, two studies applying MMC/DX/5-FU have achieved response rates of 44% and 60% in a relatively small number of patients, but those studies did not apply measurable disease as eligibility criterion, and used hardly evaluable criteria such as serum acid phosphatase levels, and sclerotic healing of lytic bone lesions, for follow-up. 120,121 The EORTC has studied MMC single agent treatment in 30 patients with measurable disease, achieving a 36% response rate. 122.

In view of this wide spectrum of antitumor activity, a widespread use may be expected. However, MMC is not a curative drug and its earlier

mentioned rare but severe side effects prevent more frequent use. For similar reasons, use in adjuvant chemotherapy regimens cannot be advocated. Obviously there is a need for analog development, aimed at retaining the antitumor effect and diminishing the side effects. In view of its unique activity, MMC-analog development warrants thorough investigation.

#### References

- Wakaki S, Marumo H, Tomioka K. Isolation of new fractions of antitumor mitomycins. *Antibiotic Chemother* 1958; 8: 228–240.
- Shirahata K, Hirayana N. Revised absolute configuration of MMC. X-ray analysis of 1-N (p-bromobenzoyl) MMC. J Am Chem Soc 1983; 105: 7199.
- 3. Underberg WJM, Lingeman H. Aspects of the chemical stability of mitomycin C and porfiromycin in acidic solution. *J Pharm Sci* 1983; 72: 549–553.
- Beynen JH, Underberg WJM. Degradation of mitomycin C in acidic solution. Int J Pharm 1985; 24: 219–229.
- 5. Stevens CL, Taylor KG, Munk ME. Chemistry and structure of mitomycin C. J Med Chem 1964; 8: 1-10.
- 6. Beynen JH, den Hartigh J, Underberg WJM. Qualitative aspects of the degradation of mitomycins in alkaline solution. *J Pharm Biomed Anal* 1985; 3: 71-79.
- Garrett ER. The physical chemical characterization of the products, equilibria and kinetics of the complex transformations of the antibiotic porfiromycin. J Med Chem 1963; 6: 488-501.
- Reddy MV, Randerath K. <sup>32</sup>P-Analysis of DNA adducts in somatic and reproductive tissues of rats treated with the anticancer antibiotic mitomycin C. *Mutat Res* 1987; 179: 75–88.
- 9. Tomasz M, Lipman R, Chowdary D., et al. Isolation and structure of a covalent cross-link adduct between mitomycin C and DNA. Science 1987; 235: 1204–1208.
- 10. Tomasz M, Lipman R. Alkylation reaction of mitomycin C at acid pH. J. Am Chem Soc 1979; 101: 6063-6067.
- Tomasz M, Lipman R, Lee MS, et al. Reaction of acidactivated mitomycin C with calf thymus DNA and model guanines: elucidation of the base-catalyzed degradation of N<sup>7</sup>-alkylguanine nucleosides. Biochemistry 1987; 26: 2010-2027.
- Lin AJ, Cosby LA, Shansky CW, et al. Potential bioreductive alkylating agents. 1. Benzoquinone derivatives. J Med Chem 1972; 15: 1247-1252.
- 13. Lin AJ, Sartorelli AC. 2,3-Dimethyl-5,6-bis (methylene)-1,4-benzoquinone. The active intermediate of bioreductive alkylating agents. J Org Chem 1973; 38: 813.
- Moore HW, Czerniak R. Naturally occurring quinones as potential bioreductive alkylating agents. Med Res Rev 1981; 1: 249-280.
- Tomasz M, Mercado CM, Olson J, et al. The mode of interaction of mitomycin C with desoxyribonucleic acid and other polynucleotides in vitro. Biochemistry 1974; 13: 4878–4887.
- Cera C, Egbertson M, Teng SP, et al. DNA cross-linking by intermediates in the mitomycin activation cascade. Biochemistry 1989; 28: 5665-5669.
- 17. Kalyanaraman B, Perez-Reyes E, Mason RP. Spin trap-

- ping and direct electron spin resonance investigations of the redox metabolism of quinone anticancer drugs. *Biochem Biophys Acta* 1980; **630**: 119–130.
- Andrews PA, Pan SS, Bachur NR. Electrochemical reductive activation of mitomycin C. J Am Chem Soc 1986; 108: 4158–4166.
- Egbertson M, Danishefsky SJ. Modeling of the electrophilic activation of mitomycins: chemical evidence for the intermediacy of a mitosene semiquinone as the active electrophile. J Am Chem Soc 1987; 109: 2204-2205.
- Kohn H, Zein N, Lin XQ, et al. Mechanistic studies on the mode of reaction of mitomycin C under catalytic and electrochemical reductive conditions. J Am Chem Soc 1987; 109: 1833–1840.
- 21. Bachur NR, Gordon SL, Gee MV. A general mechanism for microsomal activation of quinone anticancer agents to free radicals. *Cancer Res* 1978; **38**: 1745–1750.
- Lown JW, Chen HH. Evidence for the generation of free hydroxyl radicals from certain quinone antitumor antibiotics upon reductive activation in solution. *Can J Chem* 1981; 59: 390–395.
- Kennedy KA, Rockwell S, Sartorelli AC. Preferential activation of mitomycin C to cytotoxic metabolites by hypoxic tumor cells. Cancer Res 1980; 40: 2356–2360.
- Tomasz MA. H<sub>2</sub>O<sub>2</sub> generation during the redox cycle of MMC and DNA-bound MMC. Chem Biol Interact 1976; 13: 89-97.
- Bachur NR, Gordon SL, Gee MV, et al. NADPH cytochrome P-450 reductase activation of quinone anticancer agents to free radicals. Proc Natl Acad Sci USA 1979; 76: 954-957.
- Pan SS, Andrews PA, Glover CJ, et al. Reductive activation of MMC and MMC metabolites catalyzed by NADPH-cytochrome P-450 reductase and xanthine oxidase. J Biol Chem 1984; 259: 959.
- Fracasso PM, Keyes SR, Rockwell S, et al. Biotransformation of mitomycin C by NADPH-cytochrome P-450 reductase and DT-diaphorase in cultured cells. Proc AACR 1983; 24: 249.
- Keyes SR, Fracasso PM, Heimbrook DC, et al. Role of NADPH cytochrome C reductase and DT-diaphorase in the biotransformation of mitomycin C. Cancer Res 1984; 44: 5638–5643.
- Fisher JF, Olsen RA. Mechanistic aspects of mitomycin C activation by flavoprotein transhydrogenases. *Dev Bio-chemistry* 1982; 21: 240.
- Iyer VN, Szybalski W. Mitomycin and porfiromycin: chemical mechanism of activation and cross-linking of DNA. Science 1964; 145: 55-58.
- 31. Hornemann U, Iguchi K, Keller PJ, et al. Reactions of mitomycin C with potassium ethyl xanthate in neutral aqueous solution. J. Org Chem 1984; 48: 5026.
- 32. Bean M, Kohn H. Studies on the reaction of mitomycin C with potassium ethyl monothiocarbonate under reductive conditions. J Org Chem 1984; 48: 5033.
- Hashimoto Y, Shudo K, Okamoto T. Modification of deoxyribonucleic acid with reductively activated MMC. Structures of modified nucleotides. *Chem Pharm Bull* 1983; 31: 861–869.
- Tomasz M, Lipman R, Verdine G, et al. Reassignment of the guanine-binding mode of reduced mitomycin C. Biochemistry 1986; 25: 4337–4343.
- Dusre L, Covey JM, Collins C, Sinha BK. DNA damage, cytotoxicity and free radical formation by mitomycin C in human cells. Chem Biol Interactions 1989; 71: 63-78.

- Crooke ST, Bradner WT. Mitomycin C: A review. Cancer Treat Rev 1976; 3: 121-139.
- Matsumoto I, Kozaka M, Takagi Y. Analysis of the acid soluble deoxyribosidic compounds accumulated in mitomycin C treated bacteria. J Biochem Tokyo 1966; 60: 653-659.
- Orstevik J. The effect of mitomycin C on DNA synthesis in P-388 cells. Acta Path Microbiol Scand 1972; 80 (B): 729-734
- Lown JW. The molecular mechanism of action of the mitomycins. In: Carter SK, Crooke ST, eds. Mitomycin C, Current Status and New Developments New York: Academic Press, 1979: 5-26.
- Reich SD. Clinical pharmacology of mitomycin C. In: Carter SK, Crooke ST, eds. Mitomycin C, Current Status and New Developments, New York: Academic Press, 1979: 243-251
- Bayer RP, Howard Flanders P. Genetic control of DNA breakdown and repair in E. coli K-12 treated with mitomycin C or ultraviolet light. Z Vererblehre 1960; 95: 345-350.
- 42. Ishii Y, Bender M. Caffeine inhibition of prereplication repair of mitomycin C-induced DNA damage in human peripheral lymphocytes. *Mutation Res* 1978; **51**: 419–425.
- Rauth AM, Barton B, Lee CPY. Effects of caffeine on L-cells exposed to mitomycin C. Cancer Res 1970; 30: 2724-2729.
- 44. Igekami E, Amakatsu Y, Haruta M. Subcutaneous sarcomas induced by mitomycin C in mice. *Acta Phat Jap* 1972; 17: 495–501.
- 45. Marquardt O, Schmahl D, Oswald H. Experimentelle Untersuchungen uber carcinogene Wirkungen von Krebs-Chemotherapeutica und Immunosuppressiva. Ατζ-neimittel Forsch 1970; 20: 1463–1467.
- 46. Rockwell S. Effects of mitomycin C alone and in combination with X-rays on EMT-6 mouse mammary tumors in vivo. J. Natl. Cancer Inst. 1983; 71: 765-771.
- 47. Ludwig CU, Peng YM, Beaudry JN, et al. Cytotoxicity of mitomycin C on clonogenic human carcinoma cells is not enhanced by hypoxia. Cancer Chemother Pharmacol 1984; 12: 146–150.
- Sartorelli AC. The role of mitomycin antibiotics in the chemotherapy of solid tumors. *Biochem Pharmacol* 1986; 35: 67-70.
- Fracasso PM, Sartorelli AC. Cytotoxicity and DNA lesions produced by mitomycin C and porfiromycin in hypoxic and aerobic EMT-6 and Chinese hamster ovary cells. Cancer Res 1986; 46: 3939–3944.
- 50. Rockwell S. Effect of some proliferative and environmental factors on the toxicity of mitomycin C to tumor cells in vitro. Int J Cancer 1986; 38: 229-235.
- Rockwell S, Keyes SR, Sartorelli AC. Modulation of the antineoplastic efficacy of mitomycin C by dicoumarol in vivo. Cancer Chemother Pharmacol 1989; 24: 349-353.
- 52. Dulhanty AM, Li M, Whitmore G. Isolation of Chinese hamster ovary cell mutants deficient in excision repair and mitomycin C bioactivation. *Cancer Res* 1989; **49**: 117–122.
- Verweij J, den Hartigh J, Stuurman M, et al. Relationship between clinical parameters and pharmacokinetics of mitomycin C. J Cancer Res Clin Oncol 1987; 113: 91-94.
- Den Hartigh J, McVie JG, van Oort WJ, et al. Pharmacokinetics of mitomycin C in humans. Cancer Res 1983; 43: 5017–5021.
- Van Hazel GA, Scott M, Rubin J. Pharmacokinetics of mitomycin C in patients receiving the drug alone or in combination. Cancer Treat Rep 1983; 67: 805–810.

- 56. Hu E, Howell SB. Pharmacokinetics of intra-arterial mitomycin C in humans. *Cancer Res* 1983; **43**: 4474–4477.
- 57. Schilcher RB, Young JD, Ratanatharatorn V, et al. Clinical pharmacokinetics of high dose mitomycin C. Cancer Chemother Pharmacol 1984; 13: 186-198.
- 58. Buice RG, Niell HB, Sidhu P, et al. Pharmacokinetics of mitomycin C in non-oat cell carcinoma of the lung. Cancer Chemother Pharmacol 1984; 13: 1-4.
- 59. Verweij J, Stuurman M, de Vries J, et al. The difference in pharmacokinetics of mitomycin C, given either as a single agent or as a part of combination chemotherapy. J Cancer Res Clin Oncol 1986; 112: 283–284.
- Schwarz HS, Philips FS. Pharmacology of mitomycin C.
   II. Renal excretion and metabolism by tissue homogenates. J Pharmacol Exp Ther 1961; 133: 335–342.
- 61. Fujita H. Comparative studies on the blood level, tissue distribution, excretion and inactivation of anticancer drugs. *Ipn J Clin Oncol* 1971; **12**: 151-162.
- 62. Pfeifle CE, Howell SB, Ashburn WL, et al. Pharmacologic studies of intra-hepatic artery chemotherapy with degradable starch microspheres. Cancer Drug Deliver 1986; 3: 1–14.
- 63. Ensminger WE, Gyves JW, Stetson P, et al. Phase I study of hepatic arterial degradable starch microspheres and mitomycin. Cancer Res 1985; 45: 4464-4467.
- 64. Milano G, Boublil JL, Bruneton JM, et al. Systemic blood levels after intra-arterial administration of micro-encapsulated mitomycin C in cancer patients. Eur J Drug Metab Pharmacol 1985; 10: 197–201.
- Andersson M, Aronson KW, Balch, C, et al. Pharmacokinetics of intra-arterial mitomycin C with or without degradable starch microspheres (DSM) in the treatment of non-resectable liver cancer. Acta Oncol 1989; 28: 219
  222.
- 66. Adams SC, Patt YZ, Rosenblum MG. Pharmacokinetics of mitomycin C following intraperitoneal administration of MMC and floxuridine for peritoneal carcinomatosis. *Proc Am Soc Clin Oncol* 1984; **3**: 361.
- Gyves J. Pharmacology of intraperitoneal infusion of 5-fluorouracil and mitomycin C. Semin Oncol 1985; 12 (suppl 4): 29–32.
- 68. Van Oosterom AT, Jol C, de Bruyn EA, et al. Intraperitoneal chemotherapy with mitomycin C in patients with resistant ovarian cancer. In: Taguchi T, Andrysek O, eds. New Trend in Cancer Chemotherapy with Mitomycin C: Amsterdam: Excerpta Medica, 1987; 192–200.
- Wajsman Z, Dhafir RA, Pfeffer M, et al. Studies of mitomycin C absorption after intravesical treatment of superficial bladder tumors. J Urol 1984; 132: 30-33.
- Bradner WT. Oral activity of mitomycin C (NSC 26980) on Walker 256 (intramuscular) tumor. Cancer Chemother Rep 1968; 52: 389-391.
- 71. Crooke ST, Henderson M, Samson M, et al. Phase I study of oral mitomycin C. Cancer Treat Rep 1976; 60: 1633–1636.
- Van Oosterom AT, de Bruyn EA, Kuin CM, et al. Clinical and pharmacological data of mitomycin C after oral administration. Proc. 5th NCI/EORTC New Drug Symposium, 1986; Abstract 8.08.
- 73. Argenta LC, Manders EK. Mitomycin C extravasation injuries. *Cancer* 1983; **51**: 1080-1082.
- 74. Lazarus HM, Gottfried MR, Herzig RH, et al. Venoocclusive disease of the liver after high-dose mitomycin C therapy and autologous bone marrow transplantation. Cancer 1982; 49: 1789-1795.
- 75. Verweij J, van der Burg MEL, Pinedo HM. Mitomycin

- C-induced hemolytic uremic syndrome. Six case reports and review of the literature on renal, pulmonary and cardiac side effects of the drug. Radiother Oncol 1987; 8: 33–41.
- Duperray A, Tranqui L, Alix JL, Cordonnier D. Effect of mitomycin C on prostacyclin synthesis by human endothelial cells. *Biochem Pharmacol* 1988; 37: 4753–4757.
- 77. Verweij J, de Vries J, Pinedo HM. Mitomycin C-induced renal toxicity, a dose dependent side effect? Eur J Cancer Clin Oncol 1987; 23: 195–199.
- 78. Verweij J, van Zanten T, et al. Prospective study on the dose relationship of mitomycin C-induced interstitial pneumonitis. Cancer 1987; 60: 756-761.
- Verweij J, Funke-Küpper AJ, Teule GJJ, et al. A prospective study on the dose dependency of cardiotoxicity induced by mitomycin C. Med Oncol Tumor Pharmacother 1988; 5: 159–163.
- 80. Baker LH, Izbicki DO, Vaitkevicius VK. Phase II study of porfiromycin versus mitomycin-C utilizing acute intermittent schedules. *Med Ped Oncol* 1976; 2: 207–213.
- 81. Comis RL, Carter SK. Integration of chemotherapy into combined modality treatment of solid tumors. III. Gastric cancer. *Cancer Treat Rev* 1974; 1: 221–228.
- 82. Comis RL, Carter SK. A review of chemotherapy in gastric cancer. Cancer 1974; 34: 1576-1586.
- 83. Beretta G, Frashini P, Labianca R, Luporini R. The value of FAM polychemotherapy in advanced gastric carcinoma. *Proc Am Soc Clin Oncol* 1982; 23: 103.
- 84. Bitran JD, Desser RK, Kozloff MF, Billings AA, Shapiro CM. Treatment of metastatic pancreatic and gastric adenocarcinomas with 5-fluorouracil, adriamycin and mitomycin C (FAM). Cancer Treat Rep 1979; 63: 2049–2051.
- 85. Woolley PV, MacDonals JS, Smythe TA, et al. A phase II trial of ftorafur, adriamycin and mitomycin C (FAMII) in advanced gastric adenocarcinoma. Cancer 1979; 44: 1211-1214.
- Zimmerman SE, Smith FP, Schein PS. Chemotherapy of pancreatic carcinoma. Cancer 1981; 47: 1724–1728.
- 87. Aberhalden RT, Bukowski RM, Groppe CW, Hewlett JS, Weick JK. Streptozotocin (STZ) and 5-fluorouracil (5FU) with and without mitomycin-C (Mito) in the treatment of pancreatic adenocarcinoma. Proc Am Soc Clin Oncol 1977; 18: 301.
- Brown JC, Bruckner HW, Storch J, Chamberlin K, Pressman PI. Combination chemotherapy for pancreatic cancer. Proc Am Soc Clin Oncol 1980; 21: 240.
- 89. Bruckner HW, Storch JA, Brown JC, Goldberg J, Chamberlin K. Phase II trial of combination chemotherapy for pancreatic cancer with 5-fluorouracil, mitomycin C and hexamethylmelamine. *Oncology* 1983; 40: 165–169.
- Bukowski RM, Aberhalden RT, Hewlett JS. Phase II trial of streptozotocin, mitomycin C and 5-fluorouracil, in adenocarcinoma of the pancreas. Cancer Clin Trials 1980; 3: 321-324.
- 91. Oster MW, Theologides A, Cooper MR, et al. Fluorouracil + adriamycin + mitomycin (FAM) versus fluorouracil + streptozotocin + mitomycin (FSM) in advanced pancreatic cancer. Proc Am Soc Clin Oncol 1982; 1: 90.
- 92. Wiggans G, Wolley PV, MacDonald JS, Smyth TA, Ueno W, Schein PS. Phase II trials of streptozotocin, mitomycin C and 5-fluorouracil (SMF) in the treatment of advanced pancreatic cancer. Cancer 1978; 41: 387–391.
- 93. DeLena M, Brandi M, Lorusso V, Colucci G. Single agent activity of mitomycin C in breast cancer. In: Ogawa M, Rozencweig M, Staquet MJ, eds. *Mitomycin C, Current*

- Impact on Cancer Chemotherapy, Amsterdam: Excerpta Medica, 1982; 89-96.
- 94. Lopez M, Papaldo P, DiLauro L, Barduagni M, Perno CF, Barduagni A. Mitomycin C in patients with metastatic breast cancer refractory to hormone therapy and chemotherapy. Oncology 1983; 40: 244-247.
- Wise GR, Kuhn IN, Godfrey TE. Mitomycin C in large infrequent doses in breast cancer. Med Ped Oncol 1976; 2: 55-60.
- Dees A, Verweij J, van Putten WLJ, Stoter G. Mitomycin
  C is an inactive drug in the third-line treatment of
  hormone- and chemotherapy refractory breast cancer. Eur
  J Cancer Clin Oncol 1987; 23: 1343–1347.
- 97. Di Stefano A, Yap HY, Blumenstein GR. Doxorubicin, mitolactol (dibromodulcitol), and mitomycin C treatment for patients with metastatic breast cancer previously treated with cyclophosphamide, methotrexate, 5-FU, vincristine and prednisone (CMFVP). Cancer Treat Rep 1981; 65: 33-38.
- 98. Friedman MA, Marcus FS, Cassidy MJ, et al. 5-Fluorouracil + oncovin + adriamycin + mitomycin C (FOAM):
  An effective program for breast cancer, even for disease refractory to previous chemotherapy. Cancer 1983; 52: 193–197.
- Konits PM, Aisner J, van Echo PA, Lichtenfeld K, Wiernik PH. Mitomycin C and vinblastine chemotherapy for advanced breast cancer. Cancer 1981; 48: 1295–1298.
- 100. Mattson W, von Eyben F, Hallstein L, Bjelkengren G. A phase II study of combined 5-fluorouracil and mitomycin C in advanced breast cancer. Cancer 1982; 49: 217–220.
- 101. Morgan LR. Adriamycin and mitomycin C in advanced breast cancer. In: Carter SK and Crooke ST, eds. Mitomycin C, Current Status and New Developments, New York: Academic Press, 1979; 101-111.
- Rosso R, Brema F, Ardizzoni A, Conte PF, Scarsi PG, Nobile MT. Mitomycin C-5-fluorouracil combination chemotherapy for advanced breast cancer. *Proc ECCO* 1983; 2: 186.
- 103. Cohen MH, Perevodchikova NI. Single agent chemotherapy of lung cancer. In: Muggia FL, Rozencweig M, eds. Lung Cancer, Progress in Therapeutic Research, Vol 11, New York: Raven Press, 1979: 343–374.
- 104. Israel L, Chahinian P, Depierre A. Response of 65 measurable epidermoid bronchogenic tumors of known spontaneous doubling time to four different chemotherapeutic regimens. Strategic deductions. Med Ped Oncol 1975; 1: 83-93
- 105. Koons LS, Catalano RB, Harris DT. Mitomycin C in epidermoid cancer of the lung. In: Carter SK, Crooke ST, eds. Mitomycin C, Current Status and New Developments, New York: Academic Press, 1979: 189–192.
- 106. Samson MK, Comis RL, Baker LH, Ginsberg S, Fraile RJ, Crooke ST. Mitomycin C in advanced adenocarcinoma and large cell carcinoma of the lung. Cancer Treat Rep 1987; 62: 163–165.
- 107. Coates AS, Woods RL, Fox RM, Levi JA, Brodie GN, Tattersall MHN. Mitomycin C and doxorubicin in adenocarcinoma of unknown primary and non-small cell bronchogenic carcinoma. In: Ogawa M, Rozencweig M, Staquet MJ, eds. Mitomycin C, Current Impact on Cancer Chemotherapy, Amsterdam: Excerpta Medica, 1982: 113–118.
- 108. Elliott JA, Ahmedzai S, Stevenson RD, Dorward AJ, Calman KC. A randomized trial comparing vindesine (VDS) with vindesine plus cisplatinum (DDP) in operable non-small cell lung cancer. Br J Cancer 1982; 43: 485.

- 109. Fraile RJ, Samson MK, Baker LH, Talley RW. Combination chemotherapy with mitomycin C, adriamycin and cyclophosphamide in advanced adenocarcinoma and large cell carcinoma of the lung. Cancer Treat Rep 1979; 63: 1983–1987.
- 110. Miller TP, McMahon LJ, Livingston RB. Extensive adenocarcinoma and large cell undifferentiated carcinoma of the lung treated with 5-FU, vincristine and mitomycin C (FOMi). Cancer Treat Rep 1980: 64: 1241-1245.
- 111. Miller TP, Weick JK, Grozea PN, Carlin DA. Extensive adenocarcinoma and large cell undifferentiated carcinoma of the lung treated with 5-FU, vindesine and mitomycin (FEMi). Cancer Treat Rep 1982; 66: 553-556.
- 112. Rosi D, Nogeire C, Brown B, Ali M, Ewer M, Samuels M. 5-Fluorouracil, adriamycin and mitomycin (Hi-FAM) chemotherapy for adenocarcinoma of the lung. Cancer 1981; 48: 21–25.
- 113. Baker L. Study of mitomycin in cervical cancer in the United States. In: Carter SK, Crooke ST, eds. *Mitomycin C, Current Status and New Developments*, New York: Academic Press, 1979; 159–162.
- 114. Say N. Chemotherapy of pulmonary metastases from uterine cervical carcinoma. *Gan Kagakuryoho* 1982; **9**: 996.
- 115. Fluchter SH, Harzmann R, Bichler KM. Local mitomycin C therapy of transitional cell carcinoma of the bladder: serum resorption study and clinical results. In Ogawa M, Rozencweig M, Staquet MJ, eds. *Mitomycin C, Current Impact on Cancer Chemotherapy*, Amsterdam: Excerpta Medica, 1982: 143–153.

- 116. Harrison GSM, Green DF, Newling DWW, Richards B, Robinson MRG, Smith PM. A phase II study of intravesical mitomycin C in the treatment of superficial bladder cancer. *Br J Urol* 1983; **55**: 676–680.
- 117. Issels BF, Prout GR, Soloway MS, et al. Mitomycin C intravesical therapy in noninvasive bladder cancer after failure on thiotepa. Cancer 1984; 53: 1025–1028.
- 118. Soloway M. Overview of treatment of superficial bladder cancer. *Urology* 1985; **26** (suppl): 18–26.
- 119. Rozencweig M, Bleiberg H, Kenis Y. Mitomycin C therapy in advanced colorectal cancer. In: Ogawa M, Rozencweig M, Staquet MJ, eds. Mitomycin C, Current Impact on Cancer Chemotherapy, Amsterdam: Excerpta Medica, 1982: 76–88.
- 120. Kasimis BS, Moran EM, Miller JB, et al. Treatment of hormone-resistant metastatic carcinoma of the prostate with 5-FU, doxorubicin and mitomycin (FAM): A preliminary report. Cancer Treat Rep 1983; 67: 937–939.
- 121. Logothetis ChJ, Samuels ML, von Esschenbach AC, et al. Mitomycin C and 5-fluorouracil (DMF) in the treatment of metastatic hormonal refractory adenocarcinoma of the prostate, with hormonal refractory adenocarcinoma of the prostate, with a note on the staging of metastatic prostate cancer. J Clin Oncol 1983; 1: 368–379.
- 122. Jones WG, Fossa SD, Bono AV, et al. Mitomycin-C in the treatment of metastatic prostate cancer: report on an EORTC phase II study. World J Urol 1986; 4: 182–185.

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